utilized heavy-atom methods [22] and difference Fourier syntheses. Refinement was carried out as described for [Cu(2d)]BF<sub>4</sub>. Final  $R_1$  [ $F \ge 4\sigma(F)$ ] = 0.0416,  $wR_2$  [all data] = 0.100, S [ $F^2$ ] = 1.06 for 310 refined parameters. The final  $\Delta F$  synthesis showed no residual feature above 1.47 e Å<sup>-3</sup>[10e] c) Crystallographic data (excluding structure factors) for the structures reported in this paper have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication no. CCDC-111910 ([Cu(2d)]BF<sub>4</sub>) and 111911 ([Ag(2d)]BF<sub>4</sub>). Copies of the data can be obtained free of charge on application to CCDC, 12 Union Road, Cambridge CB21EZ, UK (fax: (+44) 1223-336-033; e-mail: deposit@ccdc.cam.ac.uk).

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## N-Oxidation of Epothilone A – C and O-Acyl Rearrangement to C-19- and C-21-Substituted Epothilones\*\*

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Dedicated to Professor Wolfgang Steglich on the occasion of his 65th birthday

Epothilone A (1a) and B (1b) form the parent compounds of a new family of cytotoxic and antifungal macrolides from the mycobacterium *Sorangium cellulosum*.<sup>[1]</sup> According to Bollag et al.,<sup>[2]</sup> their effect upon eucaryontic cells is based on the induction of tubulin polymerization and stabilization of microtubules, causing a massive disruption in mitosis and ultimately programmed cell death (apoptosis). This mechanism, which had been extremely rare for a long time, was first observed for taxol (paclitaxel),<sup>[3]</sup> and has recently been reported for discodermolide,<sup>[4]</sup> a polyketide from marine sponges, and eleutherobin,<sup>[5]</sup> a diterpene from soft coral. The high in vitro activity against multiresistant tumor cell lines<sup>[2, 6, 7]</sup> as well as other beneficial properties<sup>[1, 8]</sup> quickly made the epothilones interesting candidates for cancer chemotherapy and for total synthesis.<sup>[9]</sup>

First results of in vivo studies on mouse models were reported by Danishefsky et al. In these studies, not epothilone B (1b), which demonstrated the highest in vitro activity, but the significantly less active deoxyepothilone B (=epothilone D, 1d) showed a promising therapeutic range with good activity against sensitive and multiresistant tumors up to the point of complete remission.<sup>[10]</sup>

In the search for possible clinically suitable derivatives, we have concentrated our efforts on the isolation of natural structural variations<sup>[11]</sup> and the chemical modification of the epothilones obtained by fermentation.<sup>[12]</sup> We report herein the unexpectedly simple functionalization of the thiazol moiety.

During the preparation of epothilones by fermentation from *Sorangium cellulosum*, numerous structural variants from biosynthesis and biotransformation were obtained in addition to the main components A and B. Particularly

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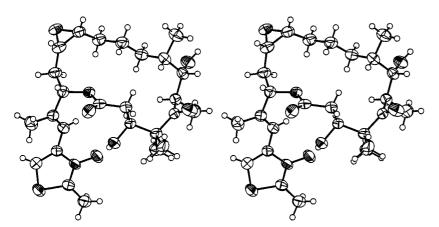
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[\*\*] This work was supported by Bristol-Myers Squibb and the Fonds der Chemischen Industrie. We thank S. Pohlan, S. Weissflog, and H. Steinmetz for their technical assistance, and Dr. V. Wray and his coworkers of the Department of Structural Research for the NMR and mass spectra. interesting are the epothilones C (1c) and D (1d), [11] as they represent the last intermediates in both the biosynthesis and several total syntheses. [9] In contrast to the biotransformation of 1c and 1d, which leads exclusively to 1a and 1b, [13] chemical epoxidation proceeds with only low chemo- and stereoselectivity. Whereas epoxidation with m-chloroperbenzoic acid at  $-20\,^{\circ}$ C leads to the formation of 1a and 1b in at least 40 and 70% yield, [14] respectively, new compounds 2a-c dominate when the same reaction is carried out at room temperature. The HPLC studies of these new compounds attracted attention as they show a band at  $\lambda_{max} = 236$  nm in the UV spectrum, which is uncharacteristic for epothilones. The main components 2a and 2b can also be obtained from the reaction of 1a and 1b with m-chloroperbenzoic acid, thus making them easily accessible in large amounts.

According to HPLC/ESI-MS and elemental analysis, 2a contains one oxygen atom more than 1a and differs only in the thiazol side chain, as can be seen in the respective  $^1H$  and  $^{13}C$  NMR spectra. As expected for a thiazol-N-oxide,  $^{[15]}$  the signals for C-18, C-19, and C-20 are shifted towards higher field by  $\Delta\delta = 9.8$  and 20. It could be confirmed by crystal structure analysis  $^{[16]}$  that an N-oxide, and not an S-oxide, was



4e H

CH<sub>3</sub>

4f H

epothilone E

epothilone F

Н

OH

TosO

Figure 1. Stereo representation of the structure of **2a** in the crystal. The heteraoatoms S, N, and O are indicated by shaded ellipsoids, and two methanol molecules are omitted for clarity.

actually formed (Figure 1).<sup>[17]</sup> The oxygen atom of the *N*-oxide is involved in strong hydrogen-bonding interactions to the OH group at C-3 (N–O 274 pm). Based on the structure of **1a**, this arrangement requires rotations around the C-1/C-2, C-2/C-3, and C-3/C-4 bonds by  $+148^{\circ}$ ,  $+31^{\circ}$ , and  $-132^{\circ}$ , [18] respectively, as well as minor changes in the torsion angles of the thiazol side chain. This conformation also exists in solution, as can be seen by the NOE effect between the axial 3-H atom and 17-H, as well as the chemical shift of the 3-OH proton at  $\delta = 6.48$  (d,  $J_{3\text{-H},3\text{-OH}} = 5.6$  Hz).

In line with rearrangements reported for  $\alpha$ -picoline and quinaldine N-oxide with carboxylic acid anhydrides, [20] we have investigated the behavior of the N-oxides **2** towards acetic anhydride. When an excess of acetic anhydride was used, a 1:1:2 mixture of **3a**, **3b**, and **4a** ( $^{1}$ H NMR) was obtained from **2a** within a few minutes at 75 °C. After 10–20 min this mixture converted completely into compound **4a** (Scheme 1). Depending on the timing of the workup, either all

Scheme 1. Reaction of the thiazol-N-oxide 2 with acetic anhydride.

three compounds or only **4a** can be obtained by silica gel chromatography. The 21-acetoxythiazol structure of **4a** can be concluded from spectroscopic data (21-CH<sub>2</sub>O:  $\delta_{\rm H}$  = 5.34 and  $\delta_{\rm C}$  = 62.5 (m)), and also from the observation that reaction with ammonia in methanol leads to epothilone E (**4f**). [11, 19]

The MS and NMR spectra reveal that  $\bf 3a$  and  $\bf 3b$  are very similar isomers of  $\bf 4a$ . Given the specific rotation values of -185.1 and 119.6,  $\bf 3a$  and  $\bf 3b$  appear to be diastereomers that differ from  $\bf 4a$  only by the presence of a 20-exo methylene group (20-CH<sub>2</sub>:  $\delta_{\rm H} = 5.2$  and 5.7,  $\delta_{\rm C} = 107$ ). This information leads to the proposed structures  $\bf I - III$ . Structure  $\bf I$ , an

expected intermediate of the total reaction, [20] can certainly be excluded since  $\bf 3a$  and  $\bf 3b$  do not undergo a change in configuration when heated to  $140\,^{\circ}{\rm C}$  in toluene. Since structure  $\bf II$ , which contains a crossconjugated chromophore, is not compatible with the observed UV spectrum ( $\lambda_{\rm max} = 267$  and  $315\,{\rm nm}$ ), only the structure of an N-acetylammonium-N-oxide ( $\bf III$ ) remains as a feasible option. However, there appears to be no precedent for this type of structure.[21]

Important for preparative applications of this *N*-oxide rearrangement is that not only other carboxylic acid anhydrides but also

activated carboxylic acid derivates such as acid chlorides can be used in the presence of 2,6-lutidine. In contrast to the case of anhydrides, the 19-chloro-, 21-chloro-, and 19-sulfonyloxy-epothilones  $\bf 4c$ ,  $\bf 4d$ , and  $\bf 4e$  were obtained with  $\it p$ -tosyl chloride and mesyl chloride/1,4-lutidine (tosyl =  $H_3CC_6H_4$ -  $SO_2$ ; mesyl =  $H_3CSO_2$ ) in dichloromethane.

Biological testing of the easily accessible thiazol derivatives of type **4** confirm earlier results,<sup>[11, 12]</sup> which suggest that structure modifications at C-21 but not at C-19 are tolerated (Table 1). The studies also reflect the trend towards higher

Table 1. Comparison of in vitro activities of the epothilone derivatives 2a-c and 4a-g with those of paclitaxel and epothilone A-D (1a-d).

Compound	L929 <sup>[a]</sup> IC <sub>50</sub> [nm]	Cell line KB-3.1 <sup>[b]</sup> IC <sub>50</sub> [nM]	А-549 <sup>[c]</sup> IС <sub>50</sub> [пм] <sup>[d]</sup>
paclitaxel (taxol)	80	12	4
epothilone A (1a)	8	2	1.4
epothilone B (1b)	1.4	1.2	0.2
epothilone C (1c)	100	40	60
epothilone D (1d)	20	24	20
2 a	200	100	160
2 b	4	2	1.5
2 c	1400	200	600
4a	30	10	5
4c	400	130	20
4d	1100	600	1100
4e	1800	1500	1500
epothilone E (4 f)	40	10	6
epothilone F (4g)	3	1.0	0.2

[a] Mouse fibroblasts (ATCC CCL1). [B] Cervix carcinoma (human; DSM ACC158). [C] Lung carcinoma (human; DSM ACC107). [D] The growth was determined by the tetrazolium reduction test.

activities of the epothilones of the B series as compared to those of the A series. For the *N*-oxides **2a** and **2b**, for instance, even a difference by a factor of 50 to 100 can be observed. Epothilone F (**4g**),<sup>[11]</sup> in vitro the most effective derivative after epothilone B, can be prepared with particular ease with the procedures presented herein starting from **1b**, which is obtained by fermentation. Furthermore, the route via the *N*-oxides **2** offers access to a variety of other derivatization and degradation reactions at C-21. It is furthermore noteworthy that the *N*-oxide rearrangement<sup>[22]</sup> which has hardly been used on thiazoles was employed for the first time on a complex natural product in the presence of sensitive functional groups.

## Experimental Section

General procedure for the preparation of the epothilone-N-oxides 2: A solution of m-chloroperbenzoic acid (2 mmol) in dichloromethane (10 mL) was added to a stirred solution of 1 (1 mmol) in dichloromethane (0.5 mL). After 3 h the reaction was quenched with an aqueous  $Na_2SO_3$  solution, and the resulting mixture distributed between an aqueous saturated  $NaHCO_3$  solution and ethyl acetate. The combined organic layers were dried over  $MgSO_4$  and concentrated in vacuo. The residue (0.6-0.7~g) was separated by chromatography using RP-18 silica gel with acetonitrile/0.05~M ammonium acetate (pH 7.5, 35/65). Yield 2a: 40~%, 2b: 48~%, 2c: 20~%.

2a: Colorless crystals from methanol; m.p.  $115\,^{\circ}$ C; TLC:  $R_{\rm f} = 0.15$  (silica gel Si60, dichloromethane/methanol (95/5), detection with vanillin/sulfuric acid, bluish gray stain upon heating to  $120\,^{\circ}$ C); HPLC:  $R_{\rm t} = 2.2$  min (HD-Sil100, C-18,  $5\,\mu$ m,  $125\,\times$ 4-mm column, acetonitrile/water (40/60),  $1.5\,$ mL min<sup>-1</sup>, detection at 254 nm); UV (MeOH):  $\lambda_{\rm max}$  ( $\epsilon$ ) = 283 sh (1700),

264 sh (4700), 236 nm (17000);  $[a]_D^{21} = -114.0$  (c = 0.9 in methanol); IR (KBr):  $\tilde{\nu}$  = 3430, 2966, 2933, 2879, 1741, 1688 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  =  $2.16 \text{ (dd, } J = 13.0, 2.6 \text{ Hz, } 2-\text{H}_a), 2.46 \text{ (t, } J = 12.0 \text{ Hz, } 2-\text{H}_b), 4.54 \text{ (br d, } J = 12.0 \text{ Hz, } 2-\text{H}_b)$ 12.0 Hz, 3-H), 6.65 (d, J = 5.6 Hz, 3-OH), 3.22 (dq, J = 2.3, 5.0 Hz, 6-H),  $3.72 \text{ (d, } J = 5.0 \text{ Hz, } 7\text{-H}), 1.26 - 1.86 \text{ (m, } 8\text{-H, } 9\text{-H}_2, 10\text{-H}_2, 11\text{-H}_2), 2.91 \text{ (m, } 3.72 \text{ ($ 12-H), 2.96 (m, 13-H), 1.66 (ddd, J = 15.2, 11.2, 9.7 Hz, 14-H<sub>a</sub>), 2.23 (brd,  $J = 15.2 \text{ Hz}, 14\text{-H}_b$ , 5.32 (d, J = 11.2 Hz, 15-H), 6.81 (br s, 17-H), 7.08 (s, 19-H), 2.59 (s,  $21-H_3$ ), 1.02 (s,  $22-H_3$ ), 1.43 (s,  $23-H_3$ ), 1.18 (d, J = 6.8 Hz,  $24-H_3$ ), 0.99 (d, J = 7.1 Hz, 25-H<sub>3</sub>), 2.07 (s, 27-H<sub>3</sub>); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  = 170.5 (C-1), 40.0 (C-2), 70.9 (C-3), 55.1 (C-4), 221.4 (C-5), 40.7 (C-6), 72.5 (C-7), 37.3 (C-8), 31.5 (C-9), 22.0 (C-10), 33.3 (C-11), 62.3(C-12), 63.0 (C-13), 33.4 (C-14), 75.6(C-15), 144.3 (C-16), 111.1 (C-17), 143.3 (C-18), 110.3 (C-19), 144.7 (C-20), 13.4 (C-21), 15.3 (C-22), 23.2 (C-23), 12.5 (C-24), 16.5 (C-25), 22.2 (C-26), 18.2 (C-27); HR-EI-MS (70 eV): *m/z* (%): 509.2483 (45,  $M^+$ ), calcd ( $C_{26}H_{39}NO_7S$ ): 509.2447; EI-MS (70 eV): m/z (%): 494 (4), 493 (4), 492 (4), 410 (48), 322 (50), 304 (18), 194 (15), 182 (52), 154 (100), 126 (44).

Preparation of **3a**, **3b**, and **4a**: a) Compound **2a** (102 mg, 0.2 mmol) was dissolved in acetic anhydride (2 mL) and heated at 75 °C for 5 min. The solution was subsequently concentrated at 30 °C/1 mbar to a viscous oil, which was separated on silica gel (Si60; hexanes/methyl *tert*-butyl ether/methanol, 66/33/1), during which first 65 mg (41 %) of **4a**, then 17 mg (11 %) each of **3a** and **3b** were eluted. b) Compound **2a** (102 mg, 0.2 mmol) was dissolved in acetic anhydride (1 mL) and heated at 75 °C for 5 min. Acetic acid (0.2 mL) was added, and the mixture was maintained at 75 °C for an additional 80 min. Workup and separation as in a) provided 81 mg (74 %) **4a**.

**3a**: Colorless oil; TLC:  $R_f = 0.66$  (for conditions see **2a**); UV (MeOH):  $\lambda_{\text{max}}$  $(\varepsilon) = 203 \ (13800), \ 267 \ (13200), \ 315 \ \text{nm} \ (5000); \ [\alpha]_{D}^{21} = -185.1 \ (c = 0.94 \ \text{in}$ CHCl<sub>3</sub>/MeOH 1:1); IR (KBr):  $\tilde{\nu} = 3446, 2965, 2936, 2877, 1742, 1691 \text{ cm}^{-1}$ ; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 2.43$  (dd, J = 14.8, 3.7 Hz, H 2<sub>a</sub>), 2.53 (dd, J = 14.8, 10.2 Hz, H  $2_b$ ), 4.13 (m, 3-H), 3.33 (d, J = 6.4 Hz, 3-OH), 1.86 (dt, J = 15.0, 7.8 Hz, 14-H<sub>a</sub>), 2.08 (m, 14-H<sub>b</sub>), 5.39 (dd, J = 7.8, 2.2 Hz, 15-H), 6.23 (br s, 17- $H), 6.95 (s, 19-H), 5.18 (s, 21-H_a), 5.71 (br s, 21-H_b), 2.26 (br s, 27-H_3), 2.12 (s, 21-H_b), 2.26 (br s, 27-H_3), 2.12 (br s,$ CH<sub>3</sub>CO); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 73.4$  (C-3), 52.8 (C-4), 151.5 (C-16), 116.0 (C-17), 158.0 (C-18), 88.7 (C-19), 166.9 (C-20), 107.2 (C-21), 20.7 (C-22), 170.2, 21.2 (acetyl); positive-ion HPLC/ESI-MS (acetonitrile/0.02 м атmonium actate buffer (pH 7)): m/z: 569 [ $M+NH_4^+$ ]. **3b**: Colorless oil; TLC:  $R_f = 0.69$  (for conditions see **2a**);  $[\alpha]_D^{21} = 119.6$  (c = 1.1 in CHCl<sub>3</sub>/ MeOH, 1/1); <sup>1</sup>H NMR (CDCl<sub>3</sub>): 1.90 (m, 14-H<sub>a</sub>), 2.09 (m, 14-H<sub>b</sub>), 5.42 (dd, J = 7.8, 2.2 Hz, 15-H), 6.92 (s, 19-H), 2.23 (s, 27-H<sub>3</sub>), 2.10 (s, CH<sub>3</sub>CO); <sup>13</sup>C NMR (CDCl<sub>3</sub>): 150.8 (C-16), 116.5 (C-17), 17.2 (C-27), 170.3, 21.0 (acetyl).

**4a**: Colorless oil; TLC:  $R_i$  = 0.45 (for conditions see **2a**); UV (MeOH):  $\lambda_{max}$  ( $\epsilon$ ) = 213 (15 800), 246 (13 500), 257 sh (9800) 275 sh (2900);  $[a]_{2}^{2l}$  = - 34.4 (c = 0.6 in methanol); IR (KBr):  $\bar{\nu}$  = 3470, 2965, 2936, 2877, 1741, 1690 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  = 2.43 (dd, J = 14.7 Hz, 3.4, 2-H<sub>a</sub>), 2.53 (dd, J = 14.7, 10.3 Hz, 2-H<sub>b</sub>), 4.13 (m, 3-H), 3.26 (d, J = 6.6 Hz, 3-OH), 1.89 (dt, J = 15.1, 8.0 Hz, 14-H<sub>a</sub>), 2.10 (ddd, J = 15.1, 4.6, 2.7 Hz, 14-H<sub>b</sub>), 5.43 (dd, J = 8.0, 2.7 Hz, 15-H), 6.60 (brs, 17-H), 7.14 (s, 19-H), 5.34 (s, 21-H<sub>2</sub>), 1.09 (s, 22-H<sub>3</sub>), 1.35 (s, 23-H<sub>3</sub>), 2.10 (s, 27-H<sub>3</sub>), 2.14 (s, Acetyl); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$  = 38.8 (C-2), 73.5 (C-3), 52.7 (C-4), 43.6 (C-6), 74.8 (C-7), 137.8 (C-16), 119.6 (C-17), 152.5 (C-18), 118 (C-19), 167.7 (C-20), 62.5 (C-21), 20.7 (C-22), 21.3 (C-23), 14.3 (C-24), 15.5 (C-27), 170.2, 20.9 (acetyl); HR-EI-MS (70 eV): m/z (%): 551.2519 (26, M<sup>+</sup>), calc (C<sub>28</sub>H<sub>41</sub>NO<sub>8</sub>S): 551.2553; EI-MS (70 eV): m/z (%): 364 (65), 224 (100), 222 (56), 209 (35), 164 (36).

**4g**: a) Compound **4b** (40 mg, 70 μmol) was dissolved in methanol (1 mL), and a 25% ammonia solution (60 μL, 8 mmol) was added. After 3 min at 40 °C the solution was concentrated to dryness in vacuo, and the residue was separated by preparative HPLC using Nucleosil C-18 with acetonitrile/water (3/7). Yield 25 mg (68%). b) Compound **2b** (40 mg, 77 μmol) was dissolved in dichloromethane (1 mL), and 2,6-lutidine (80 μL, 690 μmol) and trifluoroacetic acid anhydride (80 μL, 570 μmol) were added with stirring. After heating at 75 °C for 10 min, the solution was concentrated to dryness in vacuo. The residue was treated with 25% ammonia (0.5 mL) in THF (2 mL) and heated at 45 °C for 10 min, then concentrated to dryness in vacuo and separated by chromatography as above. Yield 31 mg (78%) colorless oil; TLC:  $R_f$  = 0.15; HPLC:  $R_t$  = 2.2 min (for conditions see **2a**); UV (MeOH):  $\lambda_{max}$  ( $\epsilon$ ) = 212 (19000), 247 nm (13 800); [ $\alpha$ ] $\frac{1}{10}$  = -27.4 ( $\epsilon$  = 0.5 in methanol); IR (KBr):  $\bar{\nu}$  = 3437, 2959, 2930, 2874, 1735, 1690 cm<sup>-1</sup>;

<sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  = 2.80 (dd, J = 7.4, 5.0 Hz, 13-H), 4.92 (s, 21-H<sub>2</sub>), 1.27 (s, 23-H<sub>3</sub>), 1.35 (s, 26-H<sub>3</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$  = 32.2 (C-11), 61.3 (C-12), 61.5 (C-13), 170.0 (C-20), 62.2 (C-21).

Received: January 21, 1999 [Z12942] German version: *Angew. Chem.* **1999**, *111*, 2090 – 2093

**Keywords:** antitumor agents • macrocycles • mycobacteria • natural products • structure – activity relationships

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- CCDC-116610. Copies of the data can be obtained free of charge on application to CCDC, 12 Union Road, Cambridge CB21EZ, UK (fax: (+44)1223-336-033; e-mail: deposit@ccdc.cam.ac.uk).
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## Molecular Beside Ionic: Crystal Structures of a 1/1 and a 1/4 Adduct of Pyridine and Formic Acid\*\*

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Dedicated to Professor Heinz Dieter Lutz on the occasion of his 65th birthday

In the context of longstanding work in this laboratory on binary adducts composed of a neutral (uncharged) selected Brønsted base and acid, two such phases have recently been identified in the system pyridine/formic acid and their crystal structures determined. With rare reported precedence in analogous systems, one is molecular and the other ionic. Phase analysis of the system has been done by differential thermal analysis (DTA) and differential scanning calorimetry (DSC) as well as temperature-dependent X-ray powder diffraction. The adducts have stoichiometries  $C_5H_5N \cdot HCOOH$  and  $C_5H_5N \cdot 4HCOOH$  and melt at 219 and 233 K, respectively. The structures have been determined at 173 and 183 K.<sup>[1]</sup>

The results are visualized in Figure 1. In the 1/1 compound, a neutral molecule of the base and one of the acid form an O–H···N hydrogen-bonded heterodimer. The 1/4 compound is a pyridinium salt, [C<sub>5</sub>H<sub>5</sub>NH][HCOO(HCOOH)<sub>3</sub>]. In the complex anion, reported here in a crystal for the first time, a central formate ion is coordinated, through rather strong O–H···O hydrogen bonds, by the three extra molecules of the acid. In these, the conformation of the carboxylic group is twice synplanar and once antiplanar. A hydrogen bond

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<sup>[\*\*]</sup> The work was supported by Fonds der Chemischen Industrie. For general assistance with diffractometry and computing, thanks are also due to Dr. Wolfgang Poll and Dr. Hartmut Wunderlich.